

RESEARCH ARTICLE OPEN ACCESS

Movement Ecology of a Coastal Foundation Seagrass Species: Insights From Genetic Data and Oceanographic Modelling

Miriam Ruocco^{1,2,3,4} | Guglielmo Lacorata^{5,6} | Luigi Palatella^{6,7} | Isabella Provera¹ | Arturo Zenone^{4,8,9} | Marco Martinez⁹ | Emanuela Dattolo^{1,4} | Jessica Pazzaglia^{1,4} | Vincenzo Maximiliano Giacalone^{4,10} | Fabio Badalamenti^{4,9} | Gabriele Procaccini^{1,4}

¹Stazione Zoologica Anton Dohrn, Napoli, Italy | ²Department of Biological, Geological and Environmental Sciences, University of Bologna, Bologna, Italy | ³Fano Marine Center, Fano, Italy | ⁴National Biodiversity Future Centre (NBFC), Palermo, Italy | ⁵CNR-ISMAR, Roma, Italy | ⁶Center of Excellence of Telesensing of the Environment and Model Prediction of Severe Events (CETEMPS), L'Aquila, Italy | ⁷Liceo Scientifico Statale "C. De Giorgi", Lecce, Italy | ⁸Stazione Zoologica Anton Dohrn, Palermo, Italy | ⁹CNR-IAS, Palermo, Italy | ¹⁰CNR-IAS, Campobello di Mazara, Italy

Correspondence: Miriam Ruocco (miriam.ruocco2@unibo.it)

Received: 17 May 2024 | Revised: 27 September 2024 | Accepted: 22 October 2024

Editor: Yunwei Dong

Funding: This work was supported by the Project Marie Hazard, Italian Ministry of University and Research (MUR) (PON03PE_00203_1).

Keywords: conservation | genetic connectivity | Lagrangian simulations | seagrass | seed dispersal | Western Sicily

ABSTRACT

Aim: Seed dispersal plays a key role in shaping the distribution and genetic complexity of seagrass populations and affects their resilience capacity under disturbance. The endemic seagrass *Posidonia oceanica* is a key component of Mediterranean coastal ecosystems, but knowledge about movement ecology in this species is limited, especially regarding seed movement pathways and dispersal potential.

Location: Western coast of Sicily (central Mediterranean).

Methods: Beach-cast fruits of the Mediterranean seagrass *P. oceanica* were collected from nine localities along the Western coast of Sicily, along with adult shoots from eight putative donor meadows. We determined pair-wise genetic differentiation between established meadows and seed cohorts. Genetic assignment tests were used to infer the most likely meadow of origin of individual seeds and were complemented with forward and backward Lagrangian simulations of dispersal.

Results: A significant genetic differentiation was found between seed pools and the most-likely meadow of origin. The genetic assignment confirmed that seeds from the same cohort originated from multiple meadows and emphasised the presence of long-distance-dispersal (LDD) events (up to hundreds of km). Genetic connectivity appeared to be greater than that predicted by oceanographic simulations, which may reflect the longer temporal scales on which gene flow is shaped, in contrast to contemporary dispersal patterns. Lagrangian simulations highlighted that fruits were physically capable of dispersing beyond the study area and that the north Tunisian coast could be a key source of propagules for the populations studied.

Main Conclusions: Our study represents a significant step forward in the understanding of *P. oceanica* movement ecology and could guide meadows' conservation and restoration actions. Our findings are significant in a broader context outside of the research area and could be the basis of similar studies in other regions, especially considering the increasing number of fruiting events recorded across the Mediterranean likely associated with ocean warming.

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2024 The Author(s). Diversity and Distributions published by John Wiley & Sons Ltd.

1 | Introduction

The movement ecology of individual organisms has a profound influence on population, community, and ecosystem dynamics over contemporary and evolutionary timescales. Also, it is associated with major current problems such as habitat fragmentation, climate change, and biological invasions (Nathan et al. 2008). For plants, unlike most animals, movement is limited to particular life-history stages, that is the dispersal of pollen or seeds, although clonal plants can spread through vegetative growth and fragmentation over small to medium distances (Levin et al. 2003). Seeds dispersal allows plants to colonise new habitats, reach sites where available resources favour regeneration, and escape pests and competition with siblings and mother (Wright et al. 2008). It also determines the spatial distribution and genetic structure of populations at local and landscape scales (Wright et al. 2008; Jahnke et al. 2016).

Despite their apparent uniformity, marine habitats are characterised by clear discontinuities, in terms of environmental conditions, water circulation patterns, geological history, and seascape features that profoundly affect species' connectivity. Indeed, these may act as dispersal barriers to gene flow, leading to genetic structuring and, eventually, isolation of populations (Evans et al. 2021). Movement patterns for most marine species are poorly characterised with respect to terrestrial systems, largely for practical reasons, such as the difficulty of tracking small propagules in large volumes of water (McMahon et al. 2014; Walther, Munguia, and Fuiman 2015). However, recent advances in ecological tools (e.g., telemetry techniques and oceanographic modelling) have allowed unprecedented insights into marine movement dynamics (Walther, Munguia, and Fuiman 2015).

Seagrasses are marine foundation species inhabiting coastal waters of most of the word's coasts, substantially contributing to key ecosystem services (Cullen-Unsworth et al. 2014; Nordlund et al. 2017). They are polyphyletic assemblages of basal monocots (order Alismatales) that reinvaded the marine environment in the early Cretaceous (Larkum, Orth, and Duarte 2006), and adapted to cope with structural and physiological challenges related to fully marine conditions (Olsen et al. 2016; Ma et al. 2024). Seagrasses exhibit two major reproductive modes, that is, vegetative and sexual reproduction. Thus, the formation of established meadows typically requires these two strategies to act in concert (Larkum, Orth, and Duarte 2006). Floating fruits represent the primary mode of seagrass dispersal into new locations, while, once seeds have settled, they typically expand by rhizomatous growth (Kendrick et al. 2012; McMahon et al. 2014). The production and dispersal of sexual propagules and the subsequent establishment of seedlings are thus critical life-history stages. Seeds' dispersal plays a key role in shaping the distribution and structure of seagrass populations (Kendrick et al. 2012; Furman et al. 2015; Ruiz-Montoya, Lowe, and Kendrick 2015; Sinclair et al. 2018). It also contributes to the genetic complexity of seagrass habitats, sustaining high overall genotypic diversity and weak genetic structuring at large spatial scales (Kendrick et al. 2017).

Due to increasing pressure from local human activities and changing climatic conditions, seagrasses are declining globally at a rapid rate (Strydom et al. 2020; Blanco-Murillo et al. 2022). Recovery of seagrasses from disturbance relies on the growth of surviving plants, as well as on the dispersal of seeds, seedlings' establishment and survival (McMahon et al. 2014). Thus, understanding the movement ecology of seagrasses would also provide a way to assess the resilience capacity of populations, including the (re)-colonisation of altered or fragmented habitats, and it could support effective management actions for conservation and restoration strategies (Kendrick et al. 2012; Evans et al. 2021; Pazzaglia et al. 2021; Provera et al. 2024). The protection of areas that are sources of sexual propagules or that receive high seed inputs, or both, should become a priority target for conservation programs (Balestri, Vallerini, and Lardicci 2017). Unfortunately, knowledge of the distribution of seagrass seeds and seedlings is scarce, and mostly based on casual observations, possibly because of the inherent difficulty in predicting where seeds will settle (Balestri, Vallerini, and Lardicci 2017).

Posidonia oceanica is one of the oldest and largest seagrass species, endemic to the Mediterranean Sea, where it forms dense meadows providing high-value ecosystem services including biodiversity support and climate change mitigation (Campagne et al. 2015; Pergent-Martini et al. 2021). Posidonia oceanica habitat has also been identified as a priority habitat under the European Commission Habitats Directive (92/43/EEC), and in several European countries, the species and/or the habitat are under specific legal protection. Flowering and recruitment (i.e., establishment of seedlings at a site) in P. oceanica have been generally considered episodic and unpredictable (Buia and Mazzella 1991; Diaz-Almela et al. 2006), and constrained by the nature of the substrate (Alagna et al. 2013, 2015; Badalamenti, Alagna, and Fici 2015). Flowers and fruits have been observed in the whole Mediterranean Sea, although with a large variability in frequency and intensity among different geographical areas as well as within the same meadows (Balestri 2004). Factors controlling flowering (and fruiting) occurrence in P. oceanica are not completely uncovered, but seem to be related to endogenous plant features, as shoot age (Balestri and Vallerini 2003; Rinaldi et al. 2023), ecological and genetic factors such as heterozygosity and relatedness (Jahnke et al. 2015). Currently, the reproductive effort of the species seems to be greater than in the past, possibly related to the strong increase in SST (Sea Surface Temperature) and in the frequency of heatwaves within the Mediterranean basin (Diaz-Almela, Marba, and Duarte 2007; Ruiz et al. 2018; Procaccini, Dattolo, and Ruocco 2023; Stipcich, La Manna, and Ceccherelli 2024). This has prompted the hypothesis that the species is reacting to climate change through an increased resource allocation to reproduction (Marín-Guirao et al. 2019).

Posidonia oceanica produces large positively buoyant fruits that may be transported hundreds of kilometres far from mother meadows under the influence of wind and surface currents before releasing seeds (Jahnke et al. 2017; Mari et al. 2020; Micheli et al. 2010; Serra et al. 2010). The seed is non-dormant, and germination occurs after its maturation inside the fruit; then, it can remain attached to the seedling for up to 2 years after germination, providing fundamental nutritional resources (Balestri et al. 2009). Investigations on the spatial patterns of seedlings' distribution at the local scale suggested a possible relationship among microhabitat type, seedling establishment, and survival

rates (Alagna et al. 2013). Despite the importance of seed dispersal in determining seagrass population structure, connectivity, and resilience, there are major gaps in our understanding of these processes in P. oceanica. Existing studies have mostly addressed realised connectivity among established meadows, based on genetic differentiation and assignment tests (Arnaud-Haond et al. 2007; Serra et al. 2010; Jahnke et al. 2017; Tutar et al. 2022), or used biophysical modelling approaches to simulate P. oceanica fruits dispersal patterns (i.e., potential connectivity) (Jahnke et al. 2017; Mari et al. 2020; Serra et al. 2010), and to identify connectivity hotspots (Mari et al. 2021). Works addressing primary source areas of sexual propagules and movement of dispersing seeds based on seed genetic assignment are almost absent. A first attempt to assess the probability of P. oceanica fruits dispersal, based on the comparison of the genetic makeup of beach-cast seeds and an adjacent meadow, was conducted in the Ligurian Sea (Italy) (Micheli et al. 2010). However, this study was limited by a small sample size and the use of lowresolution molecular markers (i.e., a few RAPD loci).

Here, we collected beach-cast P. oceanica fruits from nine localities along Western Sicily covering ~220 km of coast, along with adult shoots from eight adjacent putative donor meadows. In the study area, fruiting of P. oceanica populations has been recorded almost annually since 1997 in mid to late spring, primarily through observations of beach-cast fruits (author personal observations). Direct underwater observations are more limited due to the absence of long-term meadows' monitoring programs. All adult individuals and seeds were genotyped using 16 highly polymorphic microsatellite markers. We determined levels of genetic and genotypic richness of established meadows and seed cohorts, as well as their genetic differentiation. Our main scope was to identify key source locations of sexual propagules as well as areas that receive higher seed inputs, thus characterising the main patterns of connectivity in the study area. Genetic assignment tests to infer the meadow/s of origin of seeds and their movement pathways were complemented with forward and backward Lagrangian numerical simulations to assess all possible dynamical connections among seed sources based on surface sea currents. The coupling of genetic assignment of seeds and oceanographic connectivity modelling could provide information of critical importance for addressing the dispersal ability of this coastal foundation species, thus helping to define priority sites for conservation and/or for assisting management actions, including ecosystem restoration (Jahnke et al. 2020) to ensure persistence of healthy *P. oceanica* meadows.

2 | Methods

2.1 | Sample Collection

Posidonia oceanica fruits were collected from nine beaches (i.e., beach-cast fruits): San Nicola l'Arena (Trabia, PA)-NI s, Isola delle Femmine (PA)-IF_s, Macari (San Vito Lo Capo, TP)-MAC_s, Cornino (Custonaci, TP)-CO_s, Valderice (TP)-VA_s, Marsala (TP)-MA_s, Torre Sibiliana (TP)-SI_s, Tre Fontane (Campobello di Mazara, TP)—FO_s and San Marco (Sciacca, AG)-SM_s along the Western coast of Sicily (Figure 1, Table 1) during late spring (May-June) 2021, coinciding with the peak of fruit release of this species in the study area (Alagna et al. 2013, 2015; Provera et al. 2024; Zenone, Alagna, et al. 2020; Zenone, Filippov, et al. 2020; Zenone et al. 2022). Mature, partially open fruits containing viable seeds were selected, then seeds were immediately extracted and transported to the facilities of IAS-CNR in Castellammare Del Golfo (Sicily). Seeds were left to germinate and grow for 6 months in aquaria (30L) with continuous flow-through of natural seawater at a temperature of 20°C, salinity of 37 and under a 12:12 Light:Dark photoperiod ($80 \mu mol q m^{-2} s^{-1}$). At the end of this period, *ca*. 20 seedlings were randomly chosen for DNA extraction for each collection site (herein referred as 'seed cohorts'). The central and more mature leaves, ca. 10 cm long, were gently cleaned from epiphytes and dried with silica gel prior to DNA extraction. Along with P. oceanica fruits, we collected adult shoots from eight established shallow (~5m depth) meadows (San Nicola l'Arena-NI, Isola delle Femmine-IF, Macari-MAC, Cornino-CO, Marsala—MA, Torre Sibiliana—SI, Tre Fontane—FO and San



FIGURE 1 | Sampling sites for eight established meadows (black triangles) and nine seed cohorts (open triangles) of *Posidonia oceanica* along the Western coast of Sicily. CO, Cornino; FO, Tre Fontane; IF, Isola delle Femmine; MA, Marsala; MAC, Macari; NI, San Nicola l'Arena; SI, Torre Sibiliana; SM, San Marco; VA, Valderice.

												$uH_{ m E}$		
ocation		Latitude	Longitude	Depth	Z	MLGs	R	$N_{\rm A}$ (SD)	N_{PA}	$A_{ m R}$ (SD)	H_0 (SD)	(SD)	$F_{\rm IS}\left(p ight)$	%P
d meadows														
la l'Arena PA)	IN	38°0′56.48″ N	13°37′ 8.17″ E	8 m	16	16	1.00	3.63 (2.2)	ŝ	2.11 (0.6)	0.55 (0.2)	0.52 (0.2)	-0.068 (0.9)	100
le e (PA)	IF	38°12′18.12″ N	13°14′24.79″ E	3 m	16	15	0.93	3.13 (1.7)	ю	1.98 (0.5)	0.51 (0.3)	0.49 (0.2)	-0.063(0.8)	94
an Vito lo	MAC	38° 7'42.15″ N	12°43′35.28″ E	5-6 m	22	7	0.05	1.50(0.5)	0	1.50 (0.5)	0.47 (0.5)	0.32 (0.3)	-0.857 (1.0)	50
) aci, TP)	CO	38° 5'24.94″ N	12°39′16.20″ E	6 m	16	16	1.00	3.44 (2.0)	ю	2.15 (0.6)	0.63 (0.3)	0.54 (0.2)	-0.169(1.0)	94
(TP)	MA	37°48'48″ N	12°25′53″ E	4-5 m	19	15	0.78	3.13(1.8)	1	$1.89\ (0.5)$	0.50(0.3)	0.43 (0.2)	-0.172(1.0)	100
biliana a, TP)	SI	37°43′21.35″ N	12°28′6.20″ E	5 m	16	13	0.80	3.19 (1.7)	1	2.02 (0.6)	0.54(0.3)	0.49 (0.2)	-0.150(1.0)	100
tane bello di , TP)	FO	37°33'41.96″ N	12°42′24.80″ E	3 m	22	17	0.76	3.44 (2.1)	4	2.06 (0.6)	0.52 (0.3)	0.51 (0.2)	-0.067 (0.9)	100
rco , AG)	SM	37°29′54.88″ N	13° 0′53.45″ E	4 m	16	12	0.73	3.19 (1.6)	1	2.06 (0.5)	0.56 (0.3)	0.51 (0.2)	-0.117 (0.9)	94
orts (beach-ca	st fruits)													
ola l'Arena PA)	$\rm NI_S$	38° 0′32.64″ N	13°37'40.08″ E	I	21	21	1.00	3.81 (2.1)	7	3.73 (2.1)	0.52 (0.2)	0.50 (0.2)	-0.048 (0.8)	100
le le (PA)	IF_s	38°10′58.80″ N	13°13′59.54″ E	I	21	21	1.00	3.50 (2.3)	З	3.42 (2.2)	0.48 (0.3)	0.49 (0.2)	0.016 (0.4)	88
(San Vito lo P)	MAC_s	38° 7′32.14″ N	12°43'37.68" E	I	18	18	1.00	3.75 (2.4)	ŝ	3.74 (2.4)	0.59 (0.3)	0.52 (0.2)	-0.147 (1.0)	100
) aci, TP)	CO_s	38° 5′31.63″ N	12°39′43.16″ E	I	21	21	1.00	3.31 (1.9)	0	3.26 (1.9)	0.49 (0.2)	0.51 (0.2)	0.004 (0.5)	94
ie (TP)	VA_s	38° 4'15.72″ N	12°37′48.71″ E		20	20	1.00	3.38 (2.0)	1	3.33 (1.9)	0.53 (0.2)	0.50 (0.2)	-0.069 (0.9)	100

(Continued)	
CABLE 1	

												$uH_{ m E}$		
Sampling location		Latitude	Longitude	Depth	Z	MLGs	R	$N_{\rm A}$ (SD)	N_{PA}	$A_{ m R}$ (SD)	H_0 (SD)	(SD)	$F_{\rm IS}\left(p ight)$	$% D^{\prime \prime $
Marsala (TP)	MA_s	37°44′34.41″ N	12°28′18.57″ E	I	19	19	1.00	3.31 (2.1)	2	3.28 (2.0)	0.51(0.3)	0.48(0.2)	-0.073 (0.9)	100
Torre Sibiliana (Marsala, TP)	SI_S	37°43'25.34″ N	12°28′19.22″ E	I	21	21	1.00	4.13 (2.4)	4	3.39 (2.4)	0.48 (0.2)	0.52 (0.2)	0.067(0.1)	100
Tre Fontane (Campobello di Mazara, TP)	FO_s	37°34′18.84″ N	12°43′58.91″ E	I	20	20	1.00	3.69 (2.3)	0	3.65 (2.2)	0.56 (0.2)	0.54 (0.2)	-0.018 (0.6)	94
San Marco (Sciacca, AG)	SM_s	37°30'26.24″ N	13° 0′53.00″ E	I	20	20	1.00	3.81 (2.2)	2	3.77 (2.1)	0.55 (0.3)	0.56 (0.2)	0.025 (0.3)	100

Marco—SM) located in nearby areas (Figure 1, Table 1). These meadows were considered potential sources of the collected seeds. At each site, *ca.* 30 individual plants were collected via snorkelling at least five meters apart to minimise the risk of sampling the same genotype. Immediately after collection, leaf samples (*ca.* 10 cm long) were gently cleaned from epiphytes and dried with silica gel. As for seedlings, *ca.* 20 individuals were used for DNA extraction, for a total of 324 analysed samples (including adults and seeds).

2.2 | DNA Extraction and Microsatellite Analysis

About 60 mg of leaf tissue from seedlings and adult individuals was powdered in TissueLyser (Qiagen), and genomic DNA was isolated with the Macherey-Nagel NucleoSpin 96 Plant II kit. After isolation, DNA quality was checked by 1% agarose gel electrophoresis. All samples were genotyped at 16 microsatellite loci (Procaccini and Waycott 1998; Alberto et al. 2003; Arranz et al. 2013), assembled in two separate multiplexes and amplified by PCR using a QIAGEN Multiplex PCR Kit. Selected microsatellite regions and multiplex assembly are reported in Table S1. Genotyping was performed using an ABI Prism 3730 automated DNA sequencer (Applied Biosystems) with the PCR conditions detailed in Tutar et al. (2022). Peak identification and scoring were performed using the Peak Scanner Software 2 (Applied Biosystems).

2.3 | Genotypic and Genetic Diversity, Outlier Detection

The presence of identical multilocus genotypes (MLGs) was assessed by the software Gimlet (Valière 2002), and all the following analyses were performed only on different MLGs. For each population or seed cohort, genotypic diversity was assessed as the R ratio: R = (G-1)/(N-1), where G is the number of genotypes and N is the number of individuals (Dorken and Eckert 2001). We assessed the presence of null alleles using MicroDrop (Wang and Rosenberg 2012). Linkage disequilibrium (LD) and deviations from Hardy-Weinberg expectations (HWE) at each locus and across loci were tested with Genepop 4.7.5 (Rousset 2008), using 10,000 dememorisations, 1000 batches, and 10,000 iterations per batch. The statistical significances of LD pairwise comparisons were determined by applying the Bonferroni correction for multiple comparisons [α (0.05) divided by the number of tests]. Finally, we calculated the probability of identity (PI) in GenAlEx 6.5 (Peakall and Smouse 2012) to get an indication of the minimum number of loci needed for genetic tagging for each population or seed cohort. The mean number of alleles per locus $(N_{\rm A})$, private alleles $(N_{\rm PA})$, observed heterozygosity $(H_{\rm O})$, unbiased expected heterozygosity $(uH_{\rm F})$, and percentage of polymorphic loci (%P) were estimated with GenAlEx. Population-specific $F_{\rm IS}$ (with 1023 permutations) was calculated with Arlequin 3.5.2 (Excoffier and Lischer 2010). Mean allelic richness $(A_{\rm p})$ was calculated using the R package DiveRsity 1.9.90 (Keenan et al. 2013) using the rarefaction method to correct for variation in sample size. To identify putative outlier loci within the microsatellite set, a neutrality test was performed using two $F_{\rm ST}$ based approaches, implemented in Lositan (Antao et al. 2008) and BayeScan (Foll and Gaggiotti 2008). Lositan was run with

the following settings: 50,000 simulations under neutral mean $F_{\rm ST}$ and forced mean $F_{\rm ST}$ options, a confidence interval of 0.95 and an infinite allele model. BayeScan was used with default settings, resulting in the same probability threshold as used for Lositan. We considered as real outliers only those shared between the two methods.

2.4 | Genetic Differentiation and Population Structure

Different methods were used to determine the extent of gene flow and movement among seed cohorts in relation to the genetic structure of the established meadows. Specifically, we calculated Weir and Cockerham's F_{ST} in Arlequin, and G'_{ST} (Hedrick 2005) and D_{est} (Jost's D, Jost 2008) in GenAlEx, for pairwise comparisons, including between meadows and seed cohorts. The significance of D_{est}/G'_{ST} and F_{ST} comparisons was based on 9999 and 1000 permutations, respectively. An Analysis of Molecular Variance (AMOVA) was performed in GenAlEx with 9999 permutations to assess the portioning of variation within and between groups (established meadows and seed cohorts). A bayesian clustering analysis was then performed with Structure 2.3.4 (Pritchard, Stephens, and Donnelly 2000) for K2-K10 to identify genetic structure among established meadows and seed groups, separately, with the options admixture model, run length 100,000, 100,000 MCMC iterations, and correlated allele frequencies. Each K consisted of 10 independent runs. Structure output was estimated with Evanno ΔK (Evanno, Regnaut, and Goudet 2005) in Structure Harvester (Earl and Vonholdt 2012) and visualised with Clumpak (Kopelman et al. 2015). We also used the R package Adegenet 2.1.4 (Jombart 2008) to perform a discriminant analysis of principal components (DAPC) (Jombart and Collins 2015) for the adults and seeds' datasets, separately.

2.5 | Genetic Connectivity Between Established Meadows and Assignment of Seeds

GeneClass2 (Piry et al. 2004) was used to estimate (i) firstgeneration (F_0) migrants (FGMs), (ii) perform self-assignment of adult individuals from established meadows, and (iii) compute genetic assignment of beach-cast seeds of unknown origin. For migrant detection in established meadows (i), the statistical criterion for likelihood estimation was L_home/L_max (i.e., the ratio of the likelihood computed from the population where the individual was sampled [L_home] over the highest likelihood value among all populations sampled, including the one where the individual was collected [L_max]) (Piry et al. 2004). The partially Bayesian criterion of Rannala and Mountain (1997) was selected for likelihood computations. Associated probabilities were computed using a Monte-Carlo resampling algorithm (Paetkau et al. 2004) with 1000 permutations and a type I error of 0.05. For the self-assignment task (ii), which estimates the likelihood that an individual belongs to the population where it was sampled, the Paetkau et al. (2004) resampling algorithm was used with 100,000 simulation steps and a type I error of 0.01.

For the genetic assignment of beach-cast seeds of unknown origin (iii), the Rannala and Mountain (1997) method was selected as a criterion for computation. The probability of a meadow being a seed source was computed via a Monte-Carlo resampling algorithm (Paetkau et al. 2004) with 1,000,000 simulation steps and a type I error of 0.01. The analysis was conducted using the matrix of adult individuals from established meadows as the reference dataset after the removal of individuals identified as first-generation migrants (Jahnke et al. 2017; Underwood et al. 2007). The genetic assignment procedure is highly sensitive to genetic homogeneity. In our analysis, the level of genetic differentiation between populations ($F_{\rm ST}$ =0.076, see Section 3.2) should be sufficient to provide a reliable assignment (Christie et al. 2017).

2.6 | Lagrangian Modelling of Seed Dispersal

In order to characterise the potential (oceanographic) connectivity across sites in the study area, Lagrangian trajectories were simulated by means of ocean surface current fields provided by a general circulation model for the Mediterranean Sea (as in Jahnke et al. 2017; Tutar et al. 2022). The main idea was to evaluate particles' arrival probability, within a given time window, from a P. oceanica source to a target site. Only meadow locations could be considered for the analysis, as the model does not allow considering sites on the coastline (e.g., beaches). The numerical trajectories were let free to evolve throughout the central Mediterranean basin; thus, they could also reveal 'colonisation' patterns outside the geographical boundaries of the research area domain. The Lagrangian simulations, here presented, relied upon the Mediterranean Forecasting System (MFS; https:// medforecast.bo.ingv.it/) analysis fields for the year 2021, in the period April-June, which largely overlap the time of P. oceanica fruits collection in the study area. MFS provides daily velocity fields on a grid of 1/16° horizontal spatial resolution and 72 vertical levels (Dobricic and Pinardi 2008; Tonani et al. 2008; Oddo et al. 2009). The MFS core consists of a hydrodynamic, eddy-permitting model, with a variational data assimilation scheme, widely described elsewhere (e.g., Tonani et al. 2009; Dobricic and Pinardi 2008). To simulate the action of the missing velocity components, filtered out by the spatio-temporal resolution, a 'sub-grid' Lagrangian turbulence kinematic model was added to the numerical trajectory equations (Lacorata, Palatella, and Santoleri 2014; Lacorata and Vulpiani 2017; Lacorata et al. 2019). A key parameter for the calibration of the kinematic Lagrangian model (KLM) is the value of the mesoscale turbulent mean dissipation rate, $\varepsilon \sim O(10^{-9}) \text{ m}^2/\text{s}^3$, directly measured from Mediterranean drifter trajectories (http://www. myocean.eu) by means of dynamical system techniques, that is, the Finite-Scale Lyapunov Exponents (FSLE) (see, e.g., Corrado et al. 2017 and references therein for applications of this methodology to evaluate scale-dependent dispersion properties from surface drifter data). The KLM's role is to adjust the relative dispersion rates of the numerical trajectories as close as possible to the corresponding observational values (Lacorata, Palatella, and Santoleri 2014). This type of modelling apparatus was recently employed in various studies related to oceanographic connectivity in the Mediterranean Sea (Palatella et al. 2014; Maffucci et al. 2016; Torri et al. 2018; Falcini et al. 2020).

Both forward and backward-in-time Lagrangian numerical simulations were carried out to investigate all possible dynamic connections between sites in the study area. Backward-in-time

dispersion models can efficiently reconstruct drifters (such as seeds) trajectories by linking known arrival positions to potential sources. For the forward (direct) motion, 8000 numerical particles were released on the sea surface, in correspondence with each meadow site, and followed for a total time of 3 weeks (21 days). Valderice (VA) was excluded from the analysis since it is a collection site with no adjacent meadow. The trajectory evolution was followed over a 3-week period (21 days). This timeframe is generally considered the average lifetime of P. oceanica floating fruits (Serra et al. 2010; Jahnke et al. 2017), although experimental data are scarce. Preliminary observations revealed that when beach-cast fruits were returned to the sea and left floating within a ring of small floating buoys, seed release occurred after a maximum of 7 days (unpublished data). However, the date of detachment of the fruits from the mother plant was not known, hence the dehiscence time could be underestimated by days to weeks.

The probability that a particle, coming from a source site 'A', is recovered in a target site 'B' is assumed to be proportional to the fraction of time spent in proximity of 'B', that is, within a distance from the coast of the order of the grid step of the model, of order ~ O(1) km, in correspondence of the target site. Backward (reverse) trajectories were initialised in proximity of each target site, and the probabilities of coming from a given source site (inside or outside the system) were evaluated according to the aforementioned procedure. The choice of a large-scale ocean circulation model for the trajectory simulations does not allow a very accurate description of the coastal dynamics (Ruiz-Montoya, Lowe, and Kendrick 2015) but, on the other hand, it offers the opportunity to explore a wide domain around the study area, including open sea circulation features of the central Mediterranean basin. Further details about Lagrangian modelling methods are available in the Appendix of Supporting Information.

3 | Results

3.1 | Genetic and Clonal Diversity, Outlier Detection

Genotypic diversity (*R*) was generally high across the established meadows (R = 0.73–1), with the only exception of MAC, where only two distinct MLGs were found over 22 sampled individuals. All seed cohorts were composed of unique MLGs (R = 1) (Table 1).

Significant deviations from HWE (p < 0.05) were observed for 9 loci across adult populations [22 of 128 tests (17%)], and for 7 loci across seed cohorts [17 of 144 tests (12%)]. In the adults' dataset, we found significant linkage disequilibrium (LD) in 9 of 120 tests across all populations (8%) after applying Bonferroni correction, while the loci did not show evidence for LD in the seeds' dataset. The PI was low, ranging from 4.81E–04 in MAC to 3.82E–10 in CO, and from 4.84E–09 in VA_s to 9.49E–11 in SM_s.

Among meadows, NI, CO, and FO showed the largest number of alleles per locus (N_A) and a high % of polymorphic loci (%p = 94–100). SI_s, NI_s and SM_s were the seed cohorts with

the largest number of $N_{\rm A}$ and %p = 100% (Table 1). $N_{\rm A}$ and mean allelic richness ($A_{\rm R}$) were significantly higher ($N_{\rm A} = p < 0.01$; $A_{\rm R} = p < 0.001$, unpaired *t*-test) in seed cohorts ($N_{\rm A} = 3.63$; $A_{\rm R} = 3.51$) relative to meadows ($N_{\rm A} = 3.08$; $A_{\rm R} = 1.97$). Observed heterozygosity ($H_{\rm O}$) and mean expected heterozygosity ($uH_{\rm E}$) were similar among adult individuals ($H_{\rm O} = 0.54$; $uH_{\rm E} = 0.48$) and seeds ($H_{\rm O} = 0.52$; $uH_{\rm E} = 0.51$) (Table 1). As well, a similar number of private alleles ($N_{\rm PA}$) was present across meadows ($N_{\rm PA} = 2.0$) and seed cohorts ($N_{\rm PA} = 2.1$). *P. oceanica* meadow of FO had the largest number of private alleles ($N_{\rm PA} = 4$), while among seed groups, the highest number of $N_{\rm PA}$ was observed in SI_s ($N_{\rm PA} = 4$) (Table 1). $F_{\rm IS}$ was always negative among established meadows (from -0.857 in MAC to -0.063 in IF), whereas among seed cohorts it ranged from -0.147 in MAC_s to 0.067 in SI_s (Table 1).

The outlier analysis of adult individuals from established meadows with Lositan identified Po-4-3 as a locus under positive selection, while Pooc-044B02 and Pooc-333 were under balancing selection (Table S2). On the contrary, Lositan detected no outlier loci for the seed cohorts' dataset. BayeScan confirmed Po-4-3 as an outlier locus for the adults' dataset, while no outlier loci were identified for the seeds' dataset (Table S3). As Po-4-3 was confirmed as an outlier by two statistical approaches, we considered it a 'real' outlier and removed it from all further analyses of the adults' dataset and adult-seeds' dataset.

3.2 | Genetic Differentiation and Population Structure of Meadows and Seed Cohorts

The overall genetic distance among the established meadows $[F_{\rm ST}=0.076; G'_{ST} \text{ (Nei)}=0.095; D_{\rm est}=0.098]$ was more than the double of that detected among seed cohorts $[F_{\rm ST}=0.034; G'_{ST} \text{ (Nei)}=0.036; D_{\rm est}=0.043]$. This general pattern of differentiation was also supported by DAPC results (Figure 2a,b).

Considering all genetic distance indices, a low but significant pairwise differentiation was detected among all *P. oceanica* meadows, with few exceptions (Tables S4, S6, S8). Especially northern populations of NI, and IF formed unique clusters, while SI, MA, and SM mostly grouped together in the DAPC (Figure 2a). Among seed cohorts, there were no significant differences (neither with F_{ST} , G'_{ST} or D_{est}) between CO_s and VA_s, as well as SI_s vs. MAC_s, CO_s, and VA_s (Tables S5, S7, S9). The DAPC of seeds was largely consistent with this pattern (Figure 2b).

All seed cohorts were significantly genetically differentiated from their putative meadow of origin based on F_{ST} , G'_{ST} , and D_{est} coefficients (Tables 2, S10, S11). This provides evidence that they contain seeds from multiple sources. The structure analysis identified K=6 as the most likely number of clusters across established meadows. However, at lower Ks (K=2-3) a sub-division of northernmost (NI, IF, MAC, and CO) and southernmost populations (MA, SI, FO, and SM) was evident (Figure S1). For seed cohorts, K=2 as the most likely number of clusters (Figure S2); however, the differences in estimated probability among Ks were relatively small, and there was a strong degree of admixture between clusters. Further



FIGURE 2 | Discriminant analysis of principal components (DAPC) for the adults' (A) and seed cohorts' datasets (B) of *Posidonia oceanica* in Western Sicily. Each dot represents an individual contained in a population/cohort by a circle. CO, Cornino; FO, Tre Fontane; IF, Isola delle Femmine; MA, Marsala; MAC, Macari; NI, San Nicola l'Arena; SI, Torre Sibiliana; SM, San Marco; VA, Valderice.

sub-structuring becomes evident when assuming higher Ks (K=3-6), as the close relationship between CO_s and VA_s (Figure S2).

The AMOVA showed that the majority of variance was present within individuals (95%) (Table S12).

3.3 | Genetic Connectivity Among Meadows and Individual Assignment of Seeds

Only 63 out of 106 (59.4%) adult individuals were correctly assigned to the meadow of origin, indicating quite strong connectivity of *P. oceanica* in the study area. CO and FO had the

in Western Si	cily conside	sring only n	neutral loci.														
	IN	IF	MAC	co	MA	SI	FO	SM	NI_s	IF_s	MAC_s	CO_s	VA_s	MA_s	SI_s	FO_s	SM_s
IN		* *	*	*	* *	* * *	* *	* *	*	* *	* *	* * *	* *	* * *	*	* * *	* *
IF	0.091		*	* *	* * *	* *	* *	**	* * *	* * *	* * *	* *	* *	*	* * *	* * *	* *
MAC	0.124	0.145		*	*	* *	* *	su	*	*	* *	* *	* *	* *	*	* *	* *
CO	0.023	0.045	0.053		* * *	* *	* * *	*	* * *	* * *	* *	* * *	* *	* *	*	* *	* * *
MA	0.107	0.096	0.088	0.061		* * *	* * *	*	* * *	* * *	* * *	* * *	* * *	* * *	*	* * *	* * *
IS	0.067	0.067	0.173	0.049	0.045		* *	* * *	* * *	* * *	su	*	* * *	* *	*	* * *	* * *
FO	0.053	0.081	0.182	0.053	0.119	0.046		* * *	* * *	* * *	* * *	*	* *	* * *	*	* *	su
SM	0.067	0.068	0.091	0.026	0.020	0.042	0.056		* * *	SU	*	* *	* *	*	ns	*	* *
NI_S	0.027	0.078	0.069	0.030	0.053	0.064	0.103	0.052		* * *	* * *	* * *	* * *	* * *	* * *	* * *	* * *
IF_{-s}	0.072	0.096	0.126	0.051	0.059	0.076	0.074	0.013	0.083		* * *	* * *	* * *	* * *	*	* * *	* * *
MAC_S	0.059	0.051	0.150	0.027	0.040	0.007	0.040	0.017	0.063	0.033		*	* * *	* * *	ns	* *	* * *
CO_S	0.040	0.039	0.131	0.035	0.058	0.017	0.017	0.024	0.047	0.050	0.012		SU	* *	SU	* *	* *
VA_s	0.033	0.049	0.123	0.030	0.059	0.032	0.043	0.029	0.030	0.066	0.026	-0.003		* * *	SU	* * *	* *
MA_s	0.070	0.031	0.092	0.023	0.028	0.025	0.062	0.023	0.040	0.059	0.023	0.023	0.038		*	**	* *
SI_S	0.021	0.053	0.092	0.017	0.028	0.028	0.035	0.009	0.029	0.023	0.010	0.000	0.001	0.026		*	* *
FO_{-S}	0.072	0.052	0.102	0.036	0.052	0.030	0.029	0.018	0.068	0.041	0.018	0.018	0.030	0.022	0.014		*
SM_{-S}	0.055	0.077	0.117	0.039	0.073	0.041	0.010	0.027	0.073	0.049	0.025	0.027	0.036	0.038	0.033	0.023	
Abbreviations: * $p < 0.05$. ** $p < 0.01$.	CO, Cornino	; FO, Tre Fo	ntane; IF, Isc	ola delle Fem	mine; MA, I	Marsala; MA	ьС, Macari; N	VI, San Nicol	a l'Arena; SI	, Torre Sibilia	na; SM, San M	arco; VA, Vald	erice.				

TABLE 2 | Pairwise F_{ST} matrix (below the diagonal) and significance level (above the diagonal) based on 1023 permutations between 8 established meadows and 9 seed cohorts ('s') of *Posidonia oceanica*

largest percentage of right (self) assignment (69% and 65%, respectively), while SM had the lowest one (25%) (Table 3). GeneClass2 identified 14 significant first-generation migrants (FGMs) across meadows (13%, p < 0.05) (Table 3). These are inferred to represent individual long-distance dispersal events (LDD), with genotypes travelling a distance between a minimum of ~30 km (FO-SM) up to a maximum of ~200 km (SM-NI).

Across seed cohorts, only FO_s had the largest percentage of individual seeds (over the total n° of seeds collected at each location) assigned to the local meadow (FO, 60%) (Table 3, Figure 3). Seeds collected at all the other sites were assigned to multiple locations, including local (in bold in Table 3) and mostly non-local meadows (Table 3, Figure 3), suggesting mixing of dispersing seeds across the study area. FO, CO, MA, and SI were the meadows providing sexual propagules to the largest number of seed cohorts (i.e., key sources), including those collected hundreds of kilometres far (Table 3, Figure 4). The meadow at Tre Fontane (FO) was identified as a seed source for all locations in the study area, with varying percentages of contributed seeds (Table 3, Figure 4). The dispersal distance associated with seeds assignment between collection sites and established meadows ranged from 0.3 to 184 km, with a mean of 64 km (Table 3).

Globally, the largest percentage of seeds at each site were assigned to southernmost (S) *P. oceanica* populations (mostly FO and MA), with the only exception of NI, that shows a prevalence of seeds from northernmost (N) meadows (48% N, 43% S) and IF where the same percentage of seeds was assigned to northernmost and southernmost localities (38% N, 38% S) (Figure 3).

3.4 | Oceanographic Connectivity

The results of forward and backward-in-time simulations after 21 days of passive dispersal are presented in Figures S3, S4. Model-based oceanographic connectivity matrices for forward and backward-in-time simulations appear in Figure 5. The dark boxes appearing in the oceanographic connectivity matrices, corresponding to very unlikely row-column connections, are the signature of dynamical constraints imposed by the local marine surface circulation to the mean Lagrangian advection, that is, the preferential direction followed by the trajectories, depending on the initial conditions.

Overall, patterns of particle dispersal identified the main coastal flow going southward from NI to SM, hence the NI \rightarrow SM and SM \rightarrow NI Lagrangian pathways were not equiprobable. Particles released from NI and IF drifted to both the west and the east, but failed to reach the southernmost localities (i.e., FO and SM) (Figure S3). Also, parts of the particles were probably lost in the open sea areas and do not meet the coast in a suitable time for settlement (Figure S3). Particles originating from MAC, CO, and partially MA were drifted both northward and southward, potentially connecting all populations in the study area (Figure 5A, Figure S3). SI, FO, and SM-sourced particles mostly drifted south-east and possibly reached the Malta coast (Figure S3). Backward simulations revealed, in five cases (MAC, CO, MA, SI, and FO), significant (non-zero probability) Lagrangian connections with sources outside the study area, specifically the northern Tunisian coast, near Tunis (Figure 5B, Figure S4). This scenario is compatible with the presence of the Algerian Current which, at the entrance of the strait of Sicily, splits into three branches: one flowing eastward into the southernmost region of the Tyrrhenian Sea (responsible, in this specific case, of the long-range dynamical connection between Sicilian and Tunisian coasts) and the other two flowing through the strait, namely, the Atlantic Tunisian Current and the Atlantic Ionian Stream (Bèranger et al. 2004). As Tunisia (TUN) was recognised as an additional potential seed source, we also present forward-in-time simulations for this location in Figure S5, and TUN was included in the connectivity matrices of Figure 5. NI appeared to be among the most isolated meadows, as it could not receive seed inputs from other localities. On the contrary, FO exhibited the largest connectivity, as it could receive drifting propagules from all but NI analysed locations (Figure 5B, Figure S4).

4 | Discussion

This is the first large-scale study in which patterns of connectivity and dispersal in *P. oceanica* are investigated by means of a comparative genetic assessment of sexual propagules (seeds), along with adult shoots from established meadows. Genetic data were complemented with forward and backward Lagrangian particle tracking simulations to assess all possible dynamical connections between sources of dispersal vectors (i.e., floating fruits) based on sea surface currents. A similar study has been conducted on the congeneric species *P. australis* across the coastal waters of south-western Australia (Sinclair et al. 2018).

Posidonia oceanica is characterised by sporadic and unpredictable flowering and fruiting (Balestri 2004), with large variations occurring across regions and individual meadows (Diaz-Almela et al. 2006; André et al. 2023). Along the NW Sicilian coast, fruiting has been observed almost annually in mid to late spring and seeds collection along the Sicilian coast during late spring has been reported by several studies (Alagna et al. 2013, 2015; Zenone, Alagna, et al. 2020; Zenone, Filippov, et al. 2020; Zenone et al. 2022; Provera et al. 2024). However, no information is currently available on seed provenance and major sources of sexual propagules. These massive fruiting events occurring in this research area offer a precious opportunity to determine the main dispersal pathways and levels of genetic connectivity among meadows. This information is extremely relevant as it could guide proper conservation efforts and/or seeds-based restoration actions (e.g., Provera et al. 2024).

4.1 | Large Genetic and Genotypic Diversity of *P. oceanica* in the Study Area

The present analysis of individuals from eight Sicilian *P. oceanica* meadows, covering ~ 220 km coast from Palermo to Sciacca, revealed a high level of genetic and genotypic (clonal) diversity across the study area, as indicated by a large mean *R* value (0.76 \pm 0.30), and high levels of mean observed heterozygosity

Home location	Correctly assigned individuals (self-assignment)	N° of recent migrants	Assigned location (exclusion probability)	Distance travelled (~km)
NI	56%	3	SM (0.036)	212
			SI (0.027)	154
			FO (0.015)	184
IF	53%	1	SM (0.002)	169
MAC	50%	0	_	
СО	69%	2	MA (0.041)	51
			SM (0.026)	114
MA	60%	1	SM (0.021)	76
SI	54%	2	CO (0.022)	53
			SM (0.032)	58
FO	65%	2	CO (0.031)	87
			SI (0.029)	30
SM	25%	3	FO (0.035)	28
			FO (0.004)	28
			MA (0.045)	76
	Total	14 (13%)		
Beach-cast seeds	of unknown origin			
'Home' location	Individual assignment to	Nº total assign	ed Distance	N ^o not assigned

TABLE 3 | Results of the assignment tests of established meadows (adults) and seed cohorts ('s'), and data for the 14 detected first-generation migrants (adults) of *Posidonia oceanica* along the Western coast of Sicily obtained with GeneClass2.

'Home' location (beach)	Individual assignment to meadows (%) <i>p</i> > 0.1	N° total assigned individuals (%)	Distance travelled (~km)	N° not assigned individuals
NI_s	MA (33); NI (29); CO (19); FO (10)	19 (90%)	MA (142); NI (1); CO (101); FO (184)	2
IF_s	CO (24); MA (19); NI (14); FO (14); SI (5)	16 (76%)	CO (59); MA (98); NI (44); FO (146); SI (110)	5
MAC_s	CO (28); FO (28); SI (22); MA (6)	15 (83%)	CO (9); FO (92); SI (62); MA (48)	3
CO_s	FO (52); CO (24); SI (10)	18 (86%)	FO (87); CO (0.7); SI (53)	3
VA_s	FO (40); CO (30); MA (10); NI (5); SM (5)	18 (90%)	FO (83); CO (3); MA (39); NI (100); SM (108)	2
MA_s	CO (26); FO (26); SI (16); MA (11); NI (11); SM (5)	18 (95%)	CO (51); FO (42); SI (11); MA (9); NI (142); SM (76)	1
SI_s	FO (33); NI (19); MA (14); CO (10); SI (5)	17 (81%)	FO (30); NI (154); MA (11); CO (53); SI (0.3)	4
FO_s	FO (60); MA (10); SI (5); IF (5)	16 (80%)	FO (3); MA (42); SI (30); IF (146)	4

(Continues)

Beach-cast seeds o	f unknown origin			
'Home' location (beach)	Individual assignment to meadows (%) <i>p</i> > 0.1	N° total assigned individuals (%)	Distance travelled (~km)	N° not assigned individuals
SM_s	FO (60); CO (5); SM (5)	14 (70%)	FO (28); CO (113); SM (1.2)	6
	Total	151 (83%)		

Note: The analysis is based on only neutral loci. Travelled distance (~km) refers to the minimum possible geographic distance travelled via water between the collection site and the assigned location of origin. In bold, % seeds assigned to the local meadow.

Abbreviations: CO, Cornino; FO, Tre Fontane; IF, Isola delle Femmine; MA, Marsala; MAC, Macari; NI, San Nicola l'Arena; SI, Torre Sibiliana; SM, San Marco; VA, Valderice.



FIGURE 3 | Results of individual seed assignment tests of *Posidonia oceanica* along the Western coast of Sicily. For each collection site, the pie-chart shows % seeds, for each cohort, belonging to a putative meadow of origin based on GeneClass2 outputs. Cumulative % seeds assigned to northernmost (N=NI, IF, MAC, and CO) or southernmost (S=MA, SI, FO, and SM) meadows are also indicated. CO, Cornino; FO, Tre Fontane; IF, Isola delle Femmine; MA, Marsala; MAC, Macari; NI, San Nicola l'Arena; SI, Torre Sibiliana; SM, San Marco; VA, Valderice. Established meadows: Black triangles, seed cohorts: Open triangles. NA=not assigned.

and allelic richness ($H_0 = 0.54 \pm 0.05$; $A_R = 1.97 \pm 0.21$), with the only exception of the population from Macari (MAC), which appears to be composed by few clonal individuals, a possible misleading finding resulting from sampling a fairly large but isolated patch of P. oceanica surrounded by sand. The population of Cornino (CO) possess the largest values of clonal and genetic diversities (R = 1.0; $H_0 = 0.63$; $A_R = 2.1$). Overall, the observed levels of clonal and genetic diversity are similar to those found for P. oceanica meadows in the central Mediterranean Sea, including other Sicilian populations (R = 0.7 and $H_0 = 0.5$; Arnaud-Haond et al. 2007; Serra et al. 2010; Procaccini, Dattolo, and Ruocco 2023). Across the whole Mediterranean, P. oceanica populations in the Strait of Sicily show the highest average number of alleles and clonal diversity, in line with the previously raised hypothesis that this area could act as a transition or 'hybridization' zone between Eastern and Western Mediterranean groups (Arnaud-Haond et al. 2007; Serra et al. 2010). However, recent findings indicate comparable high levels of genetic and genotypic diversity along sectors of the Greek coasts and at the Easternmost distribution limit of the species (Litsi-Mizan et al. 2024; Tutar et al. 2022).

4.2 | Evidence of High Genetic Connectivity and Long-Range Dispersal of *P. oceanica*

In this study, we combined multiple dispersal measures to provide insights into real-time physical seed dispersal pathways (between the collection site and the inferred source meadow) and quantify both potential (through beach-cast seed assignment) and realised (through the detection of FGMs) genetic connectivity of *P. oceanica* in the study area. The analysis also spans across different temporal scales, as seed dispersal pathways based on genetic assignments reflect contemporary gene flow, while genetic distances between established meadows result from past recruitment events (Sinclair et al. 2018).

We found that beach-cast seed pools ('seed cohorts') exhibited much less genetic differentiation and structure than the established *P. oceanica* meadows. In addition, our study highlighted a significant genetic differentiation between all seed cohorts and the most likely meadow of origin (i.e., the local meadow). In support of these findings, genetic assignment tests provided evidence that such cohorts typically contain seeds from multiple



FIGURE 4 | Geographic visualisation of seed dispersal routes in *Posidonia oceanica* along the Western coast of Sicily, based on GeneClass2 assignment tests. Only seed dispersal events beyond the local meadow are displayed. Meadows that are identified as the best sources are indicated by red dots. Dispersal routes are coloured according to the source meadow (pop colours as in Figures 2 and 3), and the thickness reflects major (≥ 5 assigned seeds) or minor (< 5 assigned seeds) contribution to the seed cohort. For legibility, only the position of the established meadows is displayed on the enlarged map, except for Valderice (VA) for which the position of the collection site is shown. CO, Cornino; FO, Tre Fontane; IF, Isola delle Femmine; MA, Marsala; MAC, Macari; NI, San Nicola l'Arena; SI, Torre Sibiliana; SM, San Marco; VA, Valderice.

non-local sources, while local meadows only provide a minor contribution to the local seed pools. Similarly, a preliminary study in the Ligurian Sea found that *P. oceanica* beach-cast fruits likely originated from distant meadows, transported by currents, rather than from nearby meadows (Micheli et al. 2010). In our study, the only exception was at Tre Fontane (FO_s), where the largest proportion of seeds (60%, over the total seeds collected at that location) was assigned to the nearest meadow (FO), indicating that this meadow acts as a strong retainer (i.e., a place where released propagules can successfully remain in situ).

The genetic assignment of individual seeds, as well as the detection of first-generation migrants (FGMs) within established meadows, revealed the presence of several LDD events (*sensu* Sinclair et al. 2018), with genotypes travelling distances up to hundreds of km. However, we cannot exclude the presence of other issues, for instance, the FGMs detected in our study could also be the results of step-by-step 'migration' of genotypes, not representing real long-distance dispersal events.

The greatest seed dispersal events occurred between central and southernmost populations (CO \leftrightarrow FO), in both directions. The meadows of FO, CO, MA, and SI were identified as those providing sexual propagules to the largest number of seed cohorts (i.e., key source sites).

The northernmost populations of San Nicola (NI) and Isola delle Femmine (IF) were apparently slightly isolated from the main patterns of connectivity. However, genetic assignment tests of both adults and seeds revealed these meadows could be the target of long-distance dispersal events from other meadows in the study area. In support of this, both meadows showed among the highest levels of genetic and genotypic richness, indicative of high levels of sexual reproduction and/or recruitment of foreign genotypes. The long-distance dispersal events detected in our study (up to 200 km) were also hypothesized for other seagrass species with floating reproductive structures. These studies predicted dispersal events occurring over scales from 10 s to 100 s km (Källström et al. 2008; Tanaka et al. 2011; Nakajima et al. 2014; Ruiz-Montoya, Lowe, and Kendrick 2015; Grech et al. 2016; Sinclair et al. 2018; Triest et al. 2018). In some extreme cases, as for the two tropical seagrasses *Enhalus acoroides* and *Thalassia hemprichii*, strong currents from frequent typhoons could increase the maximum dispersal range of fruits to as much as 400 km (Lacap et al. 2002). In *Zostera noltii*, it has been hypothesized that dispersal and germination of seeds contained in spathes of floating shoots may occasionally occur and impact gene flow among populations at up to 600 km (Jahnke et al. 2016).

4.3 | Comparison of Genetic and Oceanographic Modelling of Dispersal

Lagrangian simulations of dispersal illustrated the most likely pathways particles travelled from/to established meadows based on hydrodynamic forces and provided potential for contemporary oceanographic connectivity within and beyond the study area. Our analysis highlighted that the mean coastal flow was directed southward, from NI to SM, while the opposite Lagrangian pathway (from SM to NI) was not equiprobable. This is somehow in contrast with seed dispersal routes based on genetic assignment, as southern populations appeared to provide sexual propagules to northernmost sites (although the opposite pattern was also detected). This apparent discrepancy between genetic movement pathways and the evidence of a preferential north-to-south coastal transport direction remains, at present, an open question. Although model simulations indicate that seeds originating from the southernmost meadows have a very low probability to travel upstream, a remote possibility that





FIGURE 5 | Oceanographic connectivity matrices for forward-intime (A) and backward-in-time (B) Lagrangian simulations. Colour bars represent the probability of site-to-site connections, within a 21day time interval, normalised to the sum over all columns for each row. San Nicola l'Arena—NI, Isola delle Femmine—IF, Macari—MAC, Cornino—CO, Marsala—MA, Torre Sibiliana—SI, Tre Fontane—FO, and San Marco—SM, Tunisia—TUN.

this might actually occur, due to minor coastal circulation features not accurately captured by the model, cannot be ruled out. However, both physical modelling and population genetics substantially agreed in identifying main patterns of dispersal across central-southernmost Sicilian populations, with the greatest level of potential and realised connectivity in this research area domain.

Back trajectories indicated significant Lagrangian connections with source locations beyond the study area, as the northern Tunisian coast, where *P. oceanica* meadows have been described (Serra et al. 2010; Telesca et al. 2015). These sites could play an important role as a source of propagules for many locations across the study area. This is coherent with findings by Mari et al. (2020), which described the region centred on the Strait of

Sicily as characterised by remarkable intercontinental connectivity. In particular, the Tunisian coast was identified among the top-100 *P. oceanica* connectivity hotspots in the Mediterranean (Mari et al. 2020).

In general, genetic connectivity (potential, through beach-cast seed assignment, and realised, via FGMs, which represent successful recruitment) appeared to be higher than potential oceanographic connectivity, as previously observed in P. oceanica along the northern Turkish coastline (Tutar et al. 2022). Discrepancies between genetic and oceanographic data may be explained by several factors, including the longer temporal scales on which gene flow is shaped, in contrast to contemporary sea-current dispersal dynamics. Genetic history of populations and past evolutionary events are especially important as they influence current patterns of differentiation and connectivity among populations (Serra et al. 2010; Chefaoui, Duarte, and Serrão 2017). Furthermore, it should be considered that oceanographic simulations were only produced for the year of seed collection (2021), while inter-annual fluctuations of the transport properties along the major marine currents may cause exceptional dispersal patterns due to the occurrence of extreme weather events (see, e.g., Palatella et al. 2014). Other studies comparing genetic and physical connectivity across a range of species not necessarily find a good agreement between both approaches (see, e.g., Serra et al. 2010; Johansson et al. 2015; Quigley et al. 2022; Tavares et al. 2023). For example, localised ecological conditions can influence the genetic structure of populations in a way that cannot be captured by the biophysical models (Johansson et al. 2015; Quigley et al. 2022). Generally, the two methods corroborated the findings of each other in the same aspects, while also providing unique insights, and thus a multidisciplinary biophysical-genetic approach is always recommended to best describe connectivity patterns (Quigley et al. 2022).

5 | Conclusions

Understanding which populations act as sources or sinks, and the direction of gene flow, can help to focus conservation and restoration efforts more effectively and predict how populations might respond to future environmental pressures. In this study, we identified key sources of sexual propagules in P. oceanica along the Western coast of Sicily, based on the genetic assignment of beach-cast fruits. The importance of these meadows for the overall connectivity of P. oceanica in the study area should be monitored throughout the years to confirm main seed dispersal routes and eventually propose these as special targets for conservation measures. It is worth mentioning that two Marine Protected Areas (MPAs) are present within the research area domain, the Egadi Islands MPA, which covers 54,000 ha and includes the islands of Favignana, Levanzo, Marettimo, and the MPA of Capo Gallo and Isola delle Femmine near Palermo. Understanding potential and realised connectivity could be especially important for the design of new MPAs, the managing of existing ones or the establishment of networks of MPAs, to implement largescale conservation strategies (Andrello et al. 2015; Lagabrielle et al. 2014).

As a foundation species, *P. oceanica* is of critical importance to ecosystem functions and habitat provisioning in coastal areas. Our study represents a step forward for the understanding of *P. oceanica* movement ecology in terms of species' dispersal ability and seed movement pathways. Furthermore, it complements existing studies on *P. oceanica* genetic and oceanographic connectivity across the species distribution range in the Mediterranean Sea.

Author Contributions

M.R. and G.P. conceived the ideas and designed the study. I.P., A.Z., M.M., and V.M.G. conducted the sampling activities and the seedling maintenance. M.R. conducted all the molecular laboratory work with a significant help from E.D., J.P., and I.P. M.R. conducted the genetic data analyses. G.L. and L.P. conducted the Lagrangian modelling analysis. M.R., G.P., and F.B. led the interpretation of data. M.R. led the writing of the manuscript. All authors contributed to drafting the manuscript, data interpretation and approved the final version.

Acknowledgements

The authors thank Giuseppe Di Stefano for the technical support in rearing seeds at IAS-CNR Lab. This work has been supported by the project Marine Hazard, PON03PE_00203_1, funded by the Italian Ministry of University and Research (MUR). Project funded under the National Recovery and Resilience Plan (NRRP), Mission 4 Component 2 Investment 1.4—Call for tender No. 3138 of 16 December 2021, rectified by Decree n.3175 of 18 December 2021 of Italian Ministry of University and Research funded by the European Union—NextGenerationEU; Award Number: Project code CN_00000033, Concession Decree No. 1034 of 17 June 2022 adopted by the Italian Ministry of University and Research, CUP D33C22000960007, Project title "National Biodiversity Future Center—NBFC". Open access publishing facilitated by Universita degli Studi di Bologna, as part of the Wiley - CRUI-CARE agreement.

Conflicts of Interest

The authors declare no conflicts of interest.

Data Availability Statement

All the data underlying this article are available in the article and in its Supporting Information.

Peer Review

The peer review history for this article is available at https://www.webof science.com/api/gateway/wos/peer-review/10.1111/ddi.13944.

References

Alagna, A., T. V. Fernández, G. D. Anna, C. Magliola, S. Mazzola, and F. Badalamenti. 2015. "Assessing *Posidonia oceanica* Seedling Substrate Preference: An Experimental Determination of Seedling Anchorage Success in Rocky vs. Sandy Substrates." *PLoS One* 10, no. 4: e0125321.

Alagna, A., T. V. Fernández, A. Terlizzi, and F. Badalamenti. 2013. "Influence of Microhabitat on Seedling Survival and Growth of the Mediterranean Seagrass *Posidonia oceanica* (L.) Delile." *Estuarine, Coastal and Shelf Science* 119: 119–125.

Alberto, F., L. Correia, S. Arnaud-Haond, C. Billot, C. M. Duarte, and E. Serrão. 2003. "New Microsatellite Markers for the Endemic Mediterranean Seagrass *Posidonia oceanica*." *Molecular Ecology Notes* 3: 253–255. André, S., P. Astruch, C. F. Boudouresque, et al. 2023. "The 2022 Mass Flowering of *Posidonia oceanica* in the French Mediterranean Sea: Is It Unprecedented?" *Scientific Reports of Port-Cros National Park* 37: 65–100.

Andrello, M., M. N. Jacobi, S. Manel, W. Thuiller, and D. Mouillot. 2015. "Extending Networks of Protected Areas to Optimize Connectivity and Population Growth Rate." *Ecography* 38, no. 3: 273–282.

Antao, T., A. Lopes, R. J. Lopes, A. Beja-Pereira, and G. Luikart. 2008. "LOSITAN: A Workbench to Detect Molecular Adaptation Based on a FST-Outlier Method." *BMC Bioinformatics* 9: 1–5.

Arnaud-Haond, S., M. Migliaccio, E. Diaz-Almela, et al. 2007. "Vicariance Patterns in the Mediterranean Sea: East-West Cleavage and Low Dispersal in the Endemic Seagrass *Posidonia oceanica*." *Journal of Biogeography* 34: 963–976.

Arranz, S. E., J. C. Avarre, C. Balasundaram, et al. 2013. "Permanent Genetic Resources Added to Molecular Ecology Resources Database 1 December 2012–31 January 2013." *Molecular Ecology Resources* 13: 546–549.

Badalamenti, F., A. Alagna, and S. Fici. 2015. "Evidences of Adaptive Traits to Rocky Substrates Undermine Paradigm of Habitat Preference of the Mediterranean Seagrass *Posidonia oceanica*." *Scientific Reports* 5: 8804.

Balestri, E. 2004. "Flowering of the Seagrass *Posidonia oceanica* in a North-Western Mediterranean Coastal Area: Temporal and Spatial Variations." *Marine Biology* 145: 61–68.

Balestri, E., S. Gobert, G. Lepoint, and C. Lardicci. 2009. "Seed Nutrient Content and Nutritional Status of *Posidonia oceanica* Seedlings in the Northwestern Mediterranean Sea." *Marine Ecology Progress Series* 388: 99–109.

Balestri, E., and F. Vallerini. 2003. "Interannual Variability in Flowering of *Posidonia oceanica* in the North-Western Mediterranean Sea, and Relationships Among Shoot Age and Flowering." *Botanica Marina* 46: 525–530.

Balestri, E., F. Vallerini, and C. Lardicci. 2017. "Recruitment and Patch Establishment by Seed in the Seagrass *Posidonia oceanica*: Importance and Conservation Implications." *Frontiers in Plant Science* 8: 1067.

Bèranger, K., L. Mortier, G.-P. Gasparini, L. Gervasio, M. Astraldi, and L. Crépon. 2004. "The Dynamics of the Sicily Strait: A Comprehensive Study From Observations and Models." *Deep Sea Research Part II: Topical Studies in Oceanography* 51: 411–440.

Blanco-Murillo, F., Y. Fernández-Torquemada, A. Garrote-Moreno, C. A. Sáez, and J. L. Sánchez-Lizaso. 2022. "*Posidonia oceanica* L. (Delile) Meadows Regression: Long-Term Affection May Be Induced by Multiple Impacts." *Marine Environmental Research* 174: 105557.

Buia, M. C., and L. Mazzella. 1991. "Reproductive Phenology of the Mediterranean Seagrasses *Posidonia oceanica* (L.) Delile, *Cymodocea nodosa* (Ucria) Aschers., and *Zostera noltii* Hornem." *Aquatic Botany* 40: 343–362.

Campagne, C. S., J.-M. Salles, P. Boissery, and J. Deter. 2015. "The Seagrass *Posidonia oceanica*: Ecosystem Services Identification and Economic Evaluation of Goods and Benefits." *Marine Pollution Bulletin* 97: 391–400.

Chefaoui, R. M., C. M. Duarte, and E. A. Serrão. 2017. "Palaeoclimatic Conditions in the Mediterranean Explain Genetic Diversity of Posidonia oceanica Seagrass Meadows." *Scientific Reports* 7, no. 1: 2732.

Christie, M. R., P. G. Meirmans, O. E. Gaggiotti, R. J. Toonen, and C. White. 2017. "Disentangling the Relative Merits and Disadvantages of Parentage Analysis and Assignment Tests for Inferring Population Connectivity." *ICES Journal of Marine Science* 74: 1749–1762.

Corrado, R., G. Lacorata, L. Palatella, R. Santoleri, and E. Zambianchi. 2017. "General Characteristics of Relative Dispersion in the Ocean." *Scientific Reports* 7: 46291.

Cullen-Unsworth, L. C., L. M. Nordlund, J. Paddock, S. Baker, L. J. Mckenzie, and R. K. F. Unsworth. 2014. "Seagrass Meadows Globally as a Coupled Social–Ecological System: Implications for Human Wellbeing." *Marine Pollution Bulletin* 83: 387–397.

Diaz-Almela, E., N. Marbà, E. Álvarez, E. Balestri, J. M. Ruiz-Fernández, and C. M. Duarte. 2006. "Patterns of Seagrass (*Posidonia oceanica*) Flowering in the Western Mediterranean." *Marine Biology* 148: 723–742.

Diaz-Almela, E., N. Marba, and C. M. Duarte. 2007. "Consequences of Mediterranean Warming Events in Seagrass (*Posidonia oceanica*) Flowering Records." *Global Change Biology* 13: 224–235.

Dobricic, S., and N. Pinardi. 2008. "An Oceanographic Three-Dimensional Variational Data Assimilation Scheme." *Ocean Modelling* 22: 89–105.

Dorken, M. E., and C. G. Eckert. 2001. "Severely Reduced Sexual Reproduction in Northern Populations of a Clonal Plant, Decodon Verticillatus (Lythraceae)." *Journal of Ecology* 89: 339–350.

Earl, D. A., and B. M. Vonholdt. 2012. "STRUCTURE HARVESTER: A Website and Program for Visualizing STRUCTURE Output and Implementing the Evanno Method." *Conservation Genetics Resources* 4: 359–361.

Evanno, G., S. Regnaut, and J. Goudet. 2005. "Detecting the Number of Clusters of Individuals Using the Software STRUCTURE: A Simulation Study." *Molecular Ecology* 14: 2611–2620.

Evans, R., K. M. Mcmahon, K.-J. Van Dijk, K. Dawkins, M. N. Jacobi, and A. Vikrant. 2021. "Identification of Dispersal Barriers for a Colonising Seagrass Using Seascape Genetic Analysis." *Science of the Total Environment* 763: 143052.

Excoffier, L., and H. E. L. Lischer. 2010. "Arlequin Suite ver 3.5: A New Series of Programs to Perform Population Genetics Analyses Under Linux and Windows." *Molecular Ecology Resources* 10: 564–567.

Falcini, F., R. Corrado, M. Torri, et al. 2020. "Seascape Connectivity of European Anchovy in the Central Mediterranean Sea Revealed by Weighted Lagrangian Backtracking and Bio-Energetic Modelling." *Scientific Reports* 10: 1–13.

Foll, M., and O. Gaggiotti. 2008. "A Genome-Scan Method to Identify Selected Loci Appropriate for Both Dominant and Codominant Markers: A Bayesian Perspective." *Genetics* 180: 977–993.

Furman, B. T., L. J. Jackson, E. Bricker, and B. J. Peterson. 2015. "Sexual Recruitment in *Zostera marina*: A Patch to Landscape-Scale Investigation." *Limnology and Oceanography* 60: 584–599.

Grech, A., J. Wolter, R. Coles, et al. 2016. "Spatial Patterns of Seagrass Dispersal and Settlement." *Diversity and Distributions* 22, no. 11: 1150–1162.

Hedrick, P. W. 2005. "A Standardized Genetic Differentiation Measure." *Evolution* 59: 1633–1638.

Jahnke, M., T. Alcoverro, P. S. Lavery, K. M. Mcmahon, and G. Procaccini. 2015. "Should We Sync? Seascape-Level Genetic and Ecological Factors Determine Seagrass Flowering Patterns." *Journal of Ecology* 103: 1464–1474.

Jahnke, M., R. Casagrandi, P. Melià, et al. 2017. "Potential and Realized Connectivity of the Seagrass *Posidonia oceanica* and Their Implication for Conservation." *Diversity and Distributions* 23: 1423–1434.

Jahnke, M., A. Christensen, D. Micu, et al. 2016. "Patterns and Mechanisms of Dispersal in a Keystone Seagrass Species." *Marine Environmental Research* 117: 54–62. Jahnke, M., P. O. Moksnes, J. L. Olsen, et al. 2020. "Integrating Genetics, Biophysical, and Demographic Insights Identifies Critical Sites for Seagrass Conservation." *Ecological Applications* 30, no. 6: e02121.

Johansson, M. L., F. Alberto, D. C. Reed, et al. 2015. "Seascape Drivers of *Macrocystis pyrifera* Population Genetic Structure in the Northeast Pacific." *Molecular Ecology* 24: 4866–4885.

Jombart, T. 2008. "Adegenet: A R Package for the Multivariate Analysis of Genetic Markers." *Bioinformatics* 24: 1403–1405.

Jombart, T., and C. Collins. 2015. A Tutorial for Discriminant Analysis of Principal Components (DAPC) Using Adegenet 2.0.0. London, UK: Imperial College London, MRC Centre for Outbreak Analysis and Modelling.

Jost, L. 2008. "GST and Its Relatives Do Not Measure Differentiation." *Molecular Ecology* 17: 4015–4026.

Källström, B., A. Nyqvist, P. Åberg, M. Bodin, and C. André. 2008. "Seed Rafting as a Dispersal Strategy for Eelgrass (*Zostera marina*)." *Aquatic Botany* 88, no. 2: 148–153.

Keenan, K., P. Mcginnity, T. F. Cross, W. W. Crozier, and P. A. Prodöhl. 2013. "diveRsity: An R Package for the Estimation and Exploration of Population Genetics Parameters and Their Associated Errors." *Methods in Ecology and Evolution* 4: 782–788.

Kendrick, G. A., R. J. Orth, J. Statton, et al. 2017. "Demographic and Genetic Connectivity: The Role and Consequences of Reproduction, Dispersal and Recruitment in Seagrasses." *Biological Reviews* 92: 921–938.

Kendrick, G. A., M. Waycott, T. J. B. Carruthers, et al. 2012. "The Central Role of Dispersal in the Maintenance and Persistence of Seagrass Populations." *Bioscience* 62: 56–65.

Kopelman, N. M., J. Mayzel, M. Jakobsson, N. A. Rosenberg, and I. Mayrose. 2015. "Clumpak: A Program for Identifying Clustering Modes and Packaging Population Structure Inferences Across K." *Molecular Ecology Resources* 15: 1179–1191.

Lacap, C. D. A., J. E. Vermaat, R. N. Rollon, and H. M. Nacorda. 2002. "Propagule Dispersal of the SE Asian Seagrasses *Enhalus acoroides* and *Thalassia hemprichii.*" *Marine Ecology Progress Series* 235: 75–80.

Lacorata, G., R. Corrado, F. Falcini, and R. Santoleri. 2019. "FSLE Analysis and Validation of Lagrangian Simulations Based on Satellite-Derived GlobCurrent Velocity Data." *Remote Sensing of Environment* 221: 136–143.

Lacorata, G., L. Palatella, and R. Santoleri. 2014. "Lagrangian Predictability Characteristics of an Ocean Model." *Journal of Geophysical Research: Oceans* 119: 8029–8038.

Lacorata, G., and A. Vulpiani. 2017. "Chaotic Lagrangian Models for Turbulent Relative Dispersion." *Physical Review E* 95: 043106.

Lagabrielle, E., E. Crochelet, M. Andrello, et al. 2014. "Connecting MPAs–Eight Challenges for Science and Management." *Aquatic Conservation: Marine and Freshwater Ecosystems* 24, no. S2: 94–110.

Larkum, A. W. D., R. J. Orth, and C. M. Duarte. 2006. *Seagrasses: Biology, Ecology and Conservation.* Dordrecht, the Netherlands: Springer Netherlands.

Levin, S. A., H. C. Muller-Landau, R. Nathan, and J. Chave. 2003. "The Ecology and Evolution of Seed Dispersal: A Theoretical Perspective." *Annual Review of Ecology, Evolution, and Systematics* 34: 575–604.

Litsi-Mizan, V., C. A. García-Escudero, C. S. Tsigenopoulos, K. Tsiaras, V. Gerakaris, and E. T. Apostolaki. 2024. "Unravelling the Genetic Pattern of Seagrass (*Posidonia oceanica*) Meadows in the Eastern Mediterranean Sea." *Biodiversity and Conservation* 33, no. 1: 257–280.

Ma, X., S. Vanneste, J. Chang, et al. 2024. "Seagrass Genomes Reveal Ancient Polyploidy and Adaptations to the Marine Environment." *Nature Plants* 10: 240–255.

Maffucci, F., R. Corrado, L. Palatella, et al. 2016. "Seasonal Heterogeneity of Ocean Warming: A Mortality Sink for Ectotherm Colonizers." *Scientific Reports* 6: 1–9.

Mari, L., P. Melià, S. Fraschetti, M. Gatto, and R. Casagrandi. 2020. "Spatial Patterns and Temporal Variability of Seagrass Connectivity in the Mediterranean Sea." *Diversity and Distributions* 26, no. 2: 169–182.

Mari, L., P. Melia, M. Gatto, and R. Casagrandi. 2021. "Identification of Ecological Hotspots for the Seagrass Posidonia oceanica via Metapopulation Modeling." *Frontiers in Marine Science* 8: 628976.

Marín-Guirao, L., L. Entrambasaguas, J. M. Ruiz, and G. Procaccini. 2019. "Heat-Stress Induced Flowering Can Be a Potential Adaptive Response to Ocean Warming for the Iconic Seagrass *Posidonia oceanica*." *Molecular Ecology* 28: 2486–2501.

McMahon, K., K.-J. Van Dijk, L. Ruiz-Montoya, et al. 2014. "The Movement Ecology of Seagrasses." *Proceedings of the Royal Society B: Biological Sciences* 281: 20140878.

Micheli, C., F. Spinosa, S. Aliani, G. Gasparini, A. Molcard, and A. Peirano. 2010. "Genetic Input by *Posidonia oceanica* (L.) Delile Fruits Dispersed by Currents in the Ligurian Sea." *Plant Biosystems* 144: 333–339.

Nakajima, Y., Y. U. Matsuki, C. Lian, et al. 2014. "The Kuroshio Current Influences Genetic Diversity and Population Genetic Structure of a Tropical Seagrass, Enhalus Acoroides." *Molecular Ecology* 23, no. 24: 6029–6044.

Nathan, R., W. M. Getz, E. Revilla, et al. 2008. "A Movement Ecology Paradigm for Unifying Organismal Movement Research." *Proceedings* of the National Academy of Sciences 105: 19052–19059.

Nordlund, L. M., E. W. Koch, E. B. Barbier, and J. C. Creed. 2017. "Correction: Seagrass Ecosystem Services and Their Variability Across Genera and Geographical Regions." *PLoS One* 12: e0169942.

Oddo, P., M. Adani, N. Pinardi, C. Fratianni, M. Tonani, and D. Pettenuzzo. 2009. "A Nested Atlantic-Mediterranean Sea General Circulation Model for Operational Forecasting." *Ocean Science* 5: 461–473.

Olsen, J. L., P. Rouzé, B. Verhelst, et al. 2016. "The Genome of the Seagrass *Zostera marina* Reveals Angiosperm Adaptation to the Sea." *Nature* 530: 331–335.

Paetkau, D., R. Slade, M. Burden, and A. Estoup. 2004. "Genetic Assignment Methods for the Direct, Real-Time Estimation of Migration Rate: A Simulation-Based Exploration of Accuracy and Power." *Molecular Ecology* 13: 55–65.

Palatella, L., F. Bignami, F. Falcini, G. Lacorata, A. S. Lanotte, and R. Santoleri. 2014. "Lagrangian Simulations and Interannual Variability of Anchovy Egg and Larva Dispersal in the Sicily Channel." *Journal of Geophysical Research: Oceans* 119: 1306–1323.

Pazzaglia, J., H. M. Nguyen, A. Santillán-Sarmiento, et al. 2021. "The Genetic Component of Seagrass Restoration: What We Know and the Way Forwards." *Water* 13: 829.

Peakall, R., and P. E. Smouse. 2012. "GenAlEx 6.5: Genetic Analysis in Excel. Population Genetic Software for Teaching and Research–An Update." *Bioinformatics* 28: 2537–2539.

Pergent-Martini, C., G. Pergent, B. Monnier, C.-F. Boudouresque, C. Mori, and A. Valette-Sansevin. 2021. "Contribution of *Posidonia oceanica* Meadows in the Context of Climate Change Mitigation in the Mediterranean Sea." *Marine Environmental Research* 165: 105236. Piry, S., A. Alapetite, J.-M. Cornuet, D. Paetkau, L. Baudouin, and A. Estoup. 2004. "GENECLASS2: A Software for Genetic Assignment and First-Generation Migrant Detection." *Journal of Heredity* 95: 536–539.

Pritchard, J. K., M. Stephens, and P. Donnelly. 2000. "Inference of Population Structure Using Multilocus Genotype Data." *Genetics* 155: 945–959.

Procaccini, G., E. Dattolo, and M. Ruocco. 2023. "Genetic Diversity and Connectivity in the Mediterranean Seagrass *Posidonia oceanica*: State of Art and Future Directions." *Cahiers de Biologie Marine* 64: 105–114.

Procaccini, G., and M. Waycott. 1998. "Brief Communication. Microsatellite Loci Identified in the Seagrass *Posidonia oceanica* (L.) Delile." *Journal of Heredity* 89: 562–568.

Provera, I., M. Martinez, A. Zenone, et al. 2024. "Exploring Priming Strategies to Improve Stress Resilience of *Posidonia oceanica* Seedlings." *Marine Pollution Bulletin* 200: 116057.

Quigley, C. N., M. Roughan, R. Chaput, A. G. Jeffs, and J. P. Gardner. 2022. "Combined Biophysical and Genetic Modelling Approaches Reveal New Insights into Population Connectivity of New Zealand Green-Lipped Mussels." *Frontiers in Marine Science* 9: 971209.

Rannala, B., and J. L. Mountain. 1997. "Detecting Immigration by Using Multilocus Genotypes." *Proceedings of the National Academy of Sciences* 94: 9197–9201.

Rinaldi, A., M. Martinez, F. Badalamenti, et al. 2023. "The Ontogeny-Specific Thermal Sensitivity of the Seagrass *Posidonia oceanica.*" *Frontiers in Marine Science* 10: 1183728.

Rousset, F. 2008. "genepop'007: A Complete Re-Implementation of the Genepop Software for Windows and Linux." *Molecular Ecology Resources* 8: 103–106.

Ruiz, J., L. Marín-Guirao, R. García-Muñoz, et al. 2018. "Experimental Evidence of Warming-Induced Flowering in the Mediterranean Seagrass *Posidonia oceanica.*" *Marine Pollution Bulletin* 134: 49–54.

Ruiz-Montoya, L., R. J. Lowe, and G. A. Kendrick. 2015. "Contemporary Connectivity Is Sustained by Wind-and Current-Driven Seed Dispersal Among Seagrass Meadows." *Movement Ecology* 3: 1–14.

Serra, I. A., A. M. Innocenti, G. Di Maida, et al. 2010. "Genetic Structure in the Mediterranean Seagrass *Posidonia oceanica*: Disentangling Past Vicariance Events From Contemporary Patterns of Gene Flow." *Molecular Ecology* 19: 557–568.

Sinclair, E. A., L. Ruiz-Montoya, S. L. Krauss, et al. 2018. "Seeds in Motion: Genetic Assignment and Hydrodynamic Models Demonstrate Concordant Patterns of Seagrass Dispersal." *Molecular Ecology* 27: 5019–5034.

Stipcich, P., G. La Manna, and G. Ceccherelli. 2024. "Warming-Induced Flowering and Fruiting in the Seagrass *Posidonia oceanica* and Uncertainties due to Context-Dependent Features." *Marine Biology* 171: 67.

Strydom, S., K. Murray, S. Wilson, et al. 2020. "Too Hot to Handle: Unprecedented Seagrass Death Driven by Marine Heatwave in a World Heritage Area." *Global Change Biology* 26: 3525–3538.

Tanaka, N., T. Demise, M. Ishii, Y. Shoji, and M. Nakaoka. 2011. "Genetic Structure and Gene Flow of Eelgrass *Zostera marina* Populations in Tokyo Bay, Japan: Implications for Their Restoration." *Marine Biology* 158: 871–882.

Tavares, A. I., J. Assis, P. D. Larkin, et al. 2023. "Long Range Gene Flow Beyond Predictions From Oceanographic Transport in a Tropical Marine Foundation Species." *Scientific Reports* 13, no. 1: 9112.

Telesca, L., A. Belluscio, A. Criscoli, et al. 2015. "Seagrass Meadows (*Posidonia oceanica*) Distribution and Trajectories of Change." *Scientific Reports* 5, no. 1: 12505.

Tonani, M., N. Pinardi, S. Dobricic, I. Pujol, and C. Fratianni. 2008. "A High-Resolution Free-Surface Model of the Mediterranean Sea." *Ocean Science* 4: 1–14.

Tonani, M., N. Pinardi, C. Fratianni, et al. 2009. "Mediterranean Forecasting System: Forecast and Analysis Assessment Through Skill Scores." *Ocean Science* 5: 649–660.

Torri, M., R. Corrado, F. Falcini, et al. 2018. "Planktonic Stages of Small Pelagic Fishes (*Sardinella aurita* and *Engraulis encrasicolus*) in the Central Mediterranean Sea: The Key Role of Physical Forcings and Implications for Fisheries Management." *Progress in Oceanography* 162: 25–39.

Triest, L., T. Sierens, D. Menemenlis, and T. Van der Stocken. 2018. "Inferring Connectivity Range in Submerged Aquatic Populations (Ruppia L.) Along European Coastal Lagoons From Genetic Imprint and Simulated Dispersal Trajectories." *Frontiers in Plant Science* 9: 302159.

Tutar, O., M. Ruocco, E. Dattolo, et al. 2022. "High Levels of Genetic Diversity and Population Structure in the Mediterranean Seagrass *Posidonia oceanica* at Its Easternmost Distribution Limit." *ICES Journal of Marine Science* 79, no. 8: 2286–2297.

Underwood, J. N., L. D. Smith, M. J. van Oppen, and J. P. Gilmour. 2007. "Multiple Scales of Genetic Connectivity in a Brooding Coral on Isolated Reefs Following Catastrophic Bleaching." *Molecular Ecology* 16, no. 4: 771–784.

Valière, N. 2002. "Gimlet: A Computer Program for Analysing Genetic Individual Identification Data." *Molecular Ecology Notes* 2: 377–379.

Walther, B. D., P. Munguia, and L. A. Fuiman. 2015. "Frontiers in Marine Movement Ecology: Mechanisms and Consequences of Migration and Dispersal in Marine Habitats." *Biology Letters* 11: 20150146.

Wang, C., and N. A. Rosenberg. 2012. "MicroDrop: A Program for Estimating and Correcting for Allelic Dropout in Nonreplicated Microsatellite Genotypes Version 1.01." https://web.stanford.edu/group/rosen/berglab/microdrop.html.

Wright, S. J., A. Trakhtenbrot, G. Bohrer, et al. 2008. "Understanding Strategies for Seed Dispersal by Wind Under Contrasting Atmospheric Conditions." *Proceedings of the National Academy of Sciences* 105: 19084–19089.

Zenone, A., A. Alagna, G. D'anna, et al. 2020. "Biological Adhesion in Seagrasses: The Role of Substrate Roughness in *Posidonia oceanica* (L.) Delile Seedling Anchorage via Adhesive Root Hairs." *Marine Environmental Research* 160: 105012.

Zenone, A., F. Badalamenti, A. Alagna, S. N. Gorb, and E. Infantes. 2022. "Assessing Tolerance to the Hydrodynamic Exposure of *Posidonia oceanica* Seedlings Anchored to Rocky Substrates." *Frontiers in Marine Science* 8: 788448.

Zenone, A., A. E. Filippov, A. Kovalev, F. Badalamenti, and S. N. Gorb. 2020. "Root Hair Adhesion in *Posidonia oceanica* (L.) Delile Seedlings: A Numerical Modelling Approach." *Frontiers in Mechanical Engineering* 6: 590894.

Supporting Information

Additional supporting information can be found online in the Supporting Information section.